Bio-Plex Pro Human Chemokine Assays

Quick Guide

For Use with	Instruction Manual #
Bio-Plex Pro Human Chemokine Assays	10031990

This guide can be used to prepare and run a full 1 x 96-well assay plate. For more information on a given step, refer to the complete instruction manual. New users can go to **bio-rad.com/bio-plex** and download the manual, which includes detailed instructions and a list of kit components.

IMPORTANT! Pay close attention to **vortexing**, **shaking**, and **incubation** instructions. Deviation from the protocol may result in low assay signal and assay variability.

Initial Preparation

1. Plan the plate layout.
2. Start up/warm up the Bio-Plex Multiplex Immunoassay System (**30 min**).
   - Bring the 10x wash buffer, assay buffer, and diluents to room temperature (RT). Keep the other items on ice until needed
   - Begin to thaw frozen samples
   - Prepare 1x wash buffer. Mix 10x stock by inversion to ensure all salts are in solution. Then dilute **1 part** 10x wash buffer (60 ml) with **9 parts** distilled water (540 ml)
3. Prime the wash station for a flat bottom plate.
4. Calibrate the Bio-Plex System by following the prompts in Bio-Plex Manager Software. This can be done now or during an assay incubation step.
5. After thawing samples, prepare them according to the following guidelines.

Sample Type	Diluent	Add Bovine Serum Albumin (BSA)	Recommended Sample Dilution
Serum and plasma	Sample diluent HB	None	Fourfold (1:4)
Culture media, with serum	Culture media	None	Neat to 1:10
Culture media, serum-free	Culture media	To 0.5% final	Neat to 1:10
Lavage, other fluids	Sample diluent HB	To 0.5% final	User optimized
Lysate	Sample diluent HB	To 0.5% final	User optimized (at least 1:2 for 50 to 500 µg/ml final protein)

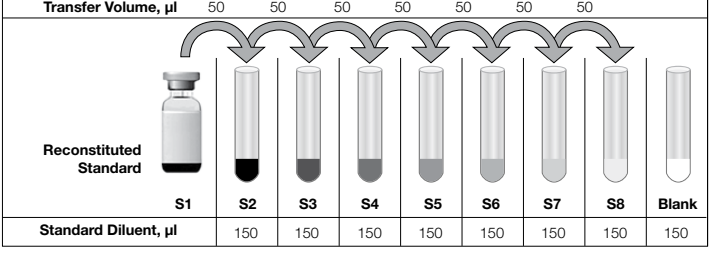
6. Reconstitute the vial of standards in **781 µl** of a diluent similar to your final sample type or matrix. Reconstitute the vial of quality controls in **250 µl** of the same diluent, as shown. **Vortex** at medium speed for **5 sec** and incubate all vials at once **on ice** for **30 min**.

Sample Type	Diluent for Standards and Controls*	Add BSA
Serum and plasma	Standard diluent HB	None
Culture media, with serum	Culture media	None
Culture media, serum-free	Culture media	To 0.5% final
Lavage, lysate, other fluids	Sample diluent HB	To 0.5% final

* If using diluents other than the standard diluent HB provided, users must establish their own control ranges.

7. Prepare a fourfold standard dilution series and blank as shown. **Vortex** at medium speed for **5 sec** between liquid transfers.

   Note: The controls are ready to use after reconstitution. No dilution is needed. Controls are included with the fixed panel only.



8. **Vortex** coupled beads at medium speed for **30 sec** and dilute to 1x in Bio-Plex Assay Buffer as shown. Protect from light.

Number of Wells	20x Beads, µl	Assay Buffer, µl	Total Volume, µl
96	288	5,472	5,760

Running the Assay

Note: Make sure all assay components are at RT before pipetting. **Vortex** at medium speed.

1. **Vortex** the diluted (1x) beads. Add **50 µl** to each well of the assay plate.
2. **Wash the plate two times** with **100 µl** Bio-Plex Wash Buffer.
3. **Vortex** samples, standards, blank, and control. Add **50 µl** to each well.
4. Cover the plate with sealing tape and protect from light with aluminum foil. Incubate on shaker at **850 ± 50 rpm** for **1 hr** at RT.
5. With 10 min left in the incubation, **vortex** detection antibodies for **15 sec** and quick-spin to collect liquid. Dilute to 1x as shown.

Number of Wells	20x Detection Antibodies, µl	Detection Antibody Diluent HB, µl	Total Volume, µl
96	150	2,850	3,000

6. **Wash the plate three times** with **100 µl** wash buffer.
7. **Vortex** the diluted (1x) detection antibodies. Add **25 µl** to each well.
8. Cover and incubate at **850 ± 50 rpm**, as in step 4, for **30 min** at RT. Meanwhile, prepare the Bio-Plex Manager Software protocol; enter standard S1 values and units provided in the assay kit.
9. With 10 min left in the incubation, **vortex** 100x streptavidin-phycoerythrin (SA-PE) for **5 sec** and quick-spin to collect liquid. Dilute to 1x as shown and protect from light.

Number of Wells	100x SA-PE, µl	Assay Buffer, µl	Total Volume, µl
96	60	5,940	6,000

10. **Wash the plate three times** with **100 µl** wash buffer.
11. **Vortex** the diluted (1x) SA-PE. Add **50 µl** to each well.

12. Cover and incubate at **850 ± 50 rpm**, as in step 4, for **10 min** at RT.

13. Wash the plate three times with **100 µl** wash buffer.

14. Resuspend the beads in **125 µl** assay buffer. Cover and shake at **850 ± 50 rpm** for **30 sec**.

15. Remove the sealing tape and **read plate** using the following settings.

Instrument	RP1 (PMT)	DD Gates	Bead Events
Bio-Plex 3D*	Standard	Select MagPlex Beads	50
Bio-Plex 200*	Low	5,000 (low); 25,000 (high)	50
Bio-Plex MAGPIX	N/A, use default instrument settings	Select MagPlex Beads	N/A

* Or similar Luminex System.

16. Quality controls are included with the fixed panel only. If they were run, then compare the observed concentrations against the ranges provided in the assay kit. Ranges apply only when standard and controls are prepared in Bio-Plex Standard Diluent HB.

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10000070643 Ver E (10031991) US/EG *19-0822 1119 Sig 0119*