EconoFit Profinity IMAC Columns, 1 and 5 ml

Instruction Manual

Catalog numbers

12009298
12009299
12009300

Please read the instructions in this manual prior to using EconoFit Profinity IMAC Columns. If you have any questions or require any further assistance, please contact your Bio-Rad Laboratories representative.

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Table of Contents

Section 1
Introduction

EconoFit Profinity IMAC Columns are convenient, disposable, prepacked low-pressure chromatography columns. EconoFit Columns offer both increased run-to-run uniformity and high purity of protein through the column design and novel resin technology. Compatible with aqueous buffers most commonly used for protein purification, EconoFit Columns offer improved performance for your protein separation needs.

Immobilized metal affinity chromatography (IMAC) is an excellent chromatography technique for purification of histidine-tagged proteins. The principle of IMAC is based on the affinity histidine has for metal ions. Side chains on the iminodiacetic acid (IDA) functional ligand selectively bind recombinant histidine-tagged proteins when the resin is charged with Ni^{2+} or other metals. The advantage of this technique is that proteins can often be purified close to homogeneity in a single step.

These columns are packed with Bio-Rad's innovative Profinity IMAC Resin. Structural characteristics such as the polymeric nature, optimized ligand density, and open pore structure of the Profinity IMAC Bead result in superb mechanical strength with high stringency, low nonspecific binding, and the ability to perform separations at high flow rates.

Section 2
Product Information

EconoFit Columns are disposable, easy-to-use, prepacked chromatographic columns supplied ready for use in convenient 1 and 5 ml sizes. They can be quickly connected to liquid chromatography systems using 10-32 fittings. Columns are available for a variety of chromatographic techniques, including desalting (size exclusion [SEC]), ion exchange (IEX), affinity (AC), mixed-mode, and hydrophobic interaction chromatography (HIC). See Table 1 for specifications. Refer to **bio-rad.com/ResinsandColumns** for a complete listing of items in the EconoFit Column product line.

Table 1. EconoFit Profinity IMAC Column specifications.

Property	Description
Size	1 and 5 ml bed volumes
Bed dimensions	1 ml: 25 mm length x 7 mm inner diameter 5 ml: 25 mm length x 16 mm inner diameter
Maximum pressure tolerance	72 psi
Recommended flow rate	1 ml: 1–2 ml/min (240–480 cm/hr) 5 ml: 5–10 ml/min (240–480 cm/hr)
Maximum flow rate	1 ml: 6 ml/min (1,440 cm/hr) 5 ml: 20 ml/min (963 cm/hr)
Fittings	10-32 (1/16"), female inlet and male outlet
Column material	Polypropylene
Frit material	High-density polyethylene
Shipping solution	20% ethanol
Storage conditions	20% ethanol
Autoclavability	Not autoclavable

Profinity IMAC Resin is also available in bottles. Refer to Ordering Information in section 10 of this manual. See Table 2 for specifications. Go to **bio-rad.com/ResinsandColumns** for more information.

Table 2. Profinity IMAC Resin specifications.

Property	Description
Functional ligand	Iminodiacetic acid (IDA)
Base bead	UNOsphere Beads
Particle size	45–90 µm
Mean particle size	60 µm
Metal ion capacity	12–30 µmol Cu^{2+}/ml
Dynamic binding capacity*	≥15 mg/ml
Recommended linear flow rate	480 cm/hr
Chemical compatibility	See Table 3
Storage temperature	4°C to ambient temperature
Storage conditions	20% ethanol
Shelf life in 20% ethanol	>1 year at ambient temperature
Operational temperature	4–40°C
Autoclavability	Not autoclavable

* Q10% determination of 1.8 mg/ml histidine6-tagged pure protein (32 kD).
Note: Dynamic binding capacity will vary from protein to protein.

Profinity IMAC Columns are compatible with the aqueous buffers most commonly used with IMAC purification techniques (see Table 3).

Table 3. Chemical compatibilities for Profinity IMAC Columns.*

Reagent	Stability
Buffer Reagents	
Tris	50 mM
HEPES	50 mM
MOPS	50 mM
Sodium or potassium phosphate	50 mM
Chelating Agent	
EDTA, EGTA	0.1 mM
Sulfhydryl Reagents	
β-mercaptoethanol	30 mM
DTT	5 mM
TCEP	10 mM
Detergents	
Nonionic	5% (Triton, Tween, NP-40)
Cationic	1% (CTAB; care must be taken to avoid protein precipitation)
Zwitterionic	5% (CHAPS, CHAPSO)
Denaturing Agents	
Guanidine HCl	6 M
Urea	8 M

continues

Reagent	Stability
Other Additives	
NaCl	2 M (include at least 300 mM NaCl in buffers)
$MgCl_2$	100 mM (use HEPES to prevent precipitation)
$CaCl_2$	5 mM (use HEPES to prevent precipitation)
Glycerol	20% (backpressure may increase significantly, slower flow rates may be required)
Ethanol	20%
Imidazole	25 mM in wash buffer, 500 mM for elution
Citrate	80 mM

* Profinity IMAC binding capacities are unaffected up to the concentrations given when employing typical reagents used for histidine-tagged protein purification.

Section 3
Buffers and Methods

IMAC methods can be run using either native or denaturing purification protocols. Under native conditions, proteins are purified using buffers that help retain the natural folded structure of the target protein. Under denaturing conditions, strong chaotropic agents (typically 6 M urea or guanidine) are added to the buffers, allowing target proteins to be purified in their unfolded states. The recommended buffer compositions and formulations are provided in Tables 4 and 5.

Table 4. Suggested buffer composition.

Step	Buffer*			
	KCl, mM	KH_2PO_4, mM	Imidazole, mM	Urea, M
Native lysis/wash buffer 1	300	50	5	NA
Native wash buffer 2	300	50	10	NA
Native elution buffer	300	50	250	NA
Denaturing lysis/wash buffer 1	300	50	5	6
Denaturing wash buffer 2	300	50	10	6
Denaturing elution buffer	300	50	250	6

* For all buffer formulations add water to 1 L, adjust pH to 8.0 with KOH or H_3PO_4, and filter through a 0.2 µm filter. Native buffers can be stored up to 1 year at 4–22°C. Denaturing buffers must be made fresh and used within 7 days or frozen in aliquots at –20°C for later use.

Table 5. Suggested buffer formulations.

Step	Buffer*			
	KCl, g	KH_2PO_4, g	Imidazole, g	Urea, g
Native lysis/wash buffer 1	22.37	6.80	0.34	NA
Native wash buffer 2	22.37	6.80	0.68	NA
Native elution buffer	22.37	6.80	17.02	NA
Denaturing lysis/wash buffer 1	22.37	6.80	0.34	360.36
Denaturing wash buffer 2	22.37	6.80	0.68	360.36
Denaturing elution buffer	22.37	6.80	17.02	360.36

* For all buffer formulations add water to 1 L, adjust pH to 8.0 with KOH or H_3PO_4 and filter through a 0.2 µm filter. Native buffers can be stored up to 1 year at 4–22°C. Denaturing buffers must be made fresh and used within 7 days or frozen in aliquots at –20°C for later use.

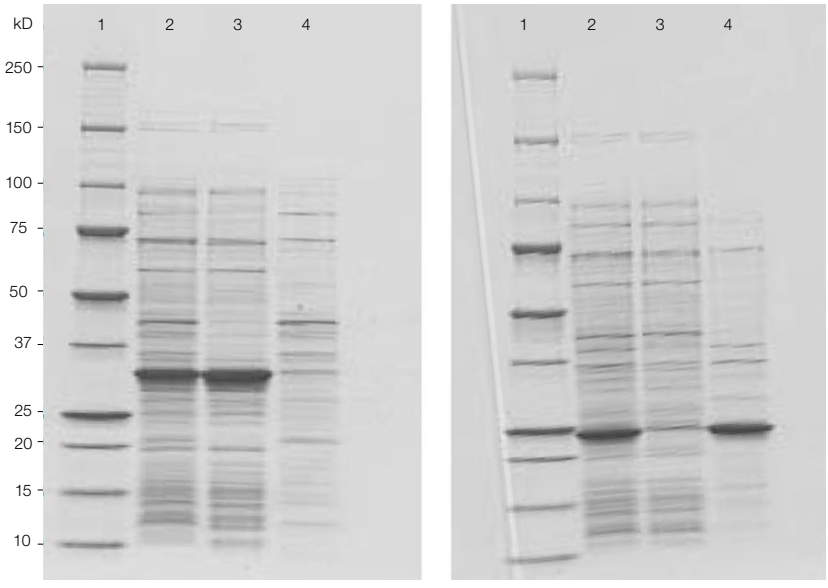
Section 4
Quick Solubility Screening Protocols

Before choosing a native or denaturing purification protocol, it is useful to determine both the approximate expression level of a protein and whether the overexpressed target protein partitions into the soluble or insoluble fraction. Soluble proteins are typically purified with the native purification procedure while insoluble proteins must be solubilized in stringent denaturants (urea or guanidine) and are purified with the denaturing procedure.

The following procedure provides a quick screen for solubility and expression level:

1. Pellet ~2 ml of *Escherichia coli* culture by centrifugation at 4,000 x g for 10 min at 4°C.
2. Resuspend the pellet in 500 µl of phosphate buffered saline (PBS) and sonicate on ice for 60 sec, in 10 sec pulses. Remove 50 µl of sonicate and label the sample Total. Centrifuge the lysate at 12,000 x g for 10 min at 4°C. Transfer the supernatant to a clean tube. Remove 50 µl of the supernatant and label the tube Soluble.
3. Resuspend the insoluble pellet in 500 µl of 6 M urea in 1x PBS and sonicate on ice for 60 sec, in 10 sec pulses. Centrifuge the lysate at 12,000 x g for 10 min at 4°C. Remove 50 µl of the supernatant and label the tube Insoluble.
4. To each of the 50 µl samples, add 150 µl of Laemmli buffer and boil for 5 min at 95°C.
5. Load 10 µl of each sample on an SDS-PAGE gel.
6. Examine the soluble and insoluble fractions for the target protein. Approximate the expression level and determine partitioning of the target protein.

A partitioning profile of soluble and insoluble target proteins, with approximate expression levels, can be seen in Figure 1.



~30% expression — soluble partitioning

~25% expression — insoluble partitioning

Fig. 1. Partitioning profiles. For both gels, Precision Plus Protein Standards, used as molecular weight markers, were loaded in lane 1, followed by the total, soluble, and insoluble fractions in lanes 2–4, respectively. The first panel depicts a 32 kD target protein, which partitions into the soluble fraction and can be purified using the native protocol (page 5). A representative chromatogram and gel for the purification of this target protein is shown in Figure 2. The second panel depicts a 24 kD target protein, which partitions into the insoluble fraction and can be purified using the denaturing protocol (section 5).

Section 5
Preparation of *E. coli* Lysates

For *E. coli* cultures expressing medium to high levels of histidine-tagged proteins (≥10% of total protein), 200 ml of culture will yield sufficient material for a 1 ml column purification, and 1,000 ml of culture will yield sufficient material for a 5 ml column purification run. For cultures expressing protein at low levels (≤10% of total protein), the culture volumes will need to be determined empirically for each protein.

Native Lysates

1. Harvest the cell pellet by centrifugation at 8,000 x g for 10 min at 4°C.
2. Determine the weight of the pellet and resuspend in 10 culture volumes of native lysis/wash buffer 1 (200 ml of culture typically yields 0.8 g of paste and results in 8 ml of lysate).
3. Sonicate the lysate on ice four times at 1 min intervals.
4. Centrifuge the lysate at 12,000 x g for 20 min at 4°C.
5. Remove the supernatant and filter it through a 0.2 µm filter immediately before applying to the column.

Denatured Lysates

1. Harvest the cell pellet by centrifugation at 8,000 x g for 10 min at 4°C.
2. Determine the weight of the pellet and resuspend in 10 culture volumes of denaturing lysis/wash buffer 1 (200 ml of culture typically yields 0.8 g of paste and results in 8 ml of lysate).
3. Sonicate the lysate four times at 1 min intervals.
4. Centrifuge the lysate at 12,000 x g for 20 min at 4°C.
5. Remove the supernatant and filter it through a 0.2 µm filter immediately before applying to the column.

Section 6
Preparing a Column and Subsequent Purification

Prepare buffer sets for either the native or denaturing purification protocols using a single buffer set throughout the procedure. To prepare the column for the procedure, remove the top closure and connect the column to the chromatography system. Open the bottom closure and connect the column outlet to the system. Flush the packing solution (20% ethanol) from the column by running two column volumes (CV) of water at a flow rate of 2 ml/min (1 ml column) or 10 ml/min (5 ml column). The column is now ready for the purification steps. Flow rates are given in ml/min and are specific to the 1 ml column.

If using a 5 ml column for a procedure, substitute the higher flow rate in the method (see Table 6).

Table 6. Purification method suggestions.

Step	Column Volumes	1 ml Column Flow Rate, ml/min	5 ml Column Flow Rate, ml/min
Equilibrate	5	2	10
Lysate load	5–10	2	10
Wash 1	6	2	10
Wash 2	6	2	10
Elute	5	2	10

Standard methods compatible with any type of chromatography system are listed in the following steps. To maximize binding capacity with large proteins (>100 kD), for purification at 4°C, or for purifications under denaturing conditions, the lysate load flow rate can be decreased (to 0.5 ml/min for the 1 ml column and 2 ml/min for the 5 ml). Whether this decrease maximizes flow rate will have to be determined empirically for individual proteins.

1. Equilibrate the column with 5 CV of equilibration/wash buffer 1 at 2 ml/min.
2. Load the sample lysate at 2 ml/min.
3. Wash the column with 6 CV of wash buffer 1 at 2 ml/min.
4. Wash the column with 6 CV of wash buffer 2 at 2 ml/min.
5. Elute the purified protein with 10 CV of elution buffer at 2 ml/min.
6. Prior to quantitation of the protein concentration, the purified protein should be exchanged into a nonimidazole buffer (imidazole can absorb at 280 nm). Purified protein from denaturing purifications should be exchanged into another buffer through dialysis.

The chromatogram and gel in Figure 2 illustrate a representative purification of a high-expressing soluble protein purified using the native buffer set and method described in Table 6.

Note: IMAC buffers made with potassium salts are more stable than sodium salt–based buffers. However, potassium will complex with sodium dodecyl sulfate (SDS) in Laemmli buffer and precipitate out of solution. Prior to analyzing IMAC samples on gels, the samples must be diluted at least 1:7 with Laemmli buffer to prevent precipitation.

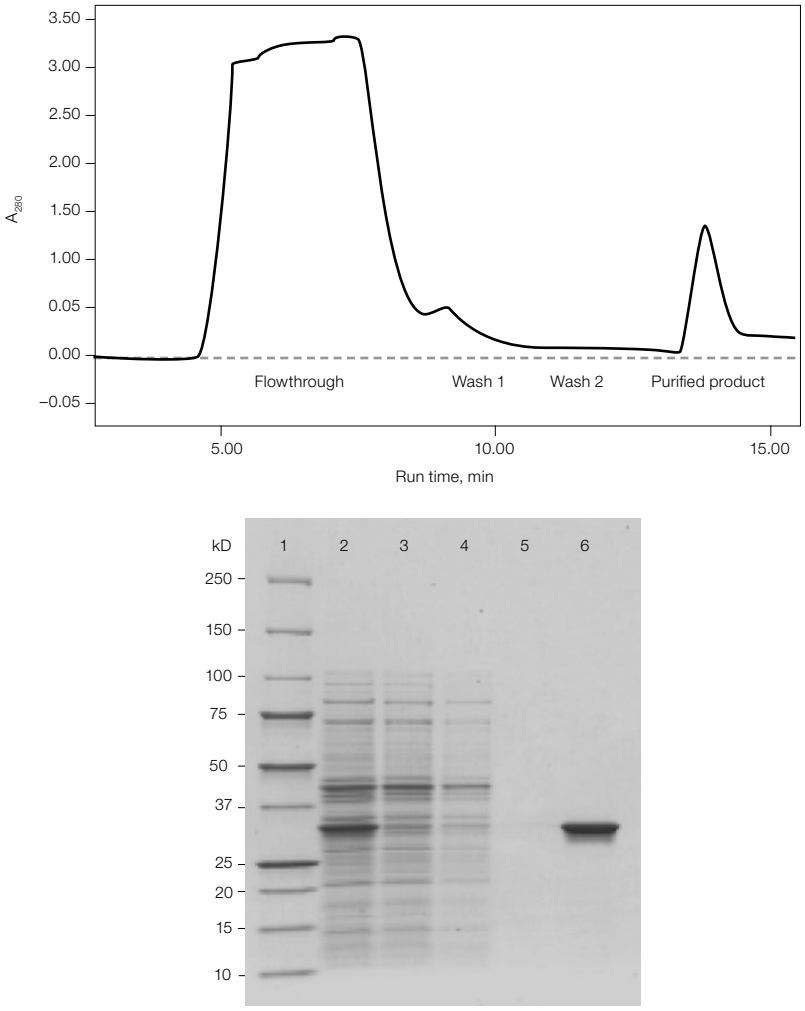


Fig. 2. Native IMAC purification. A 32 kD Nif-3* histidine-tagged protein was purified from the soluble fraction using the standard Profinity IMAC Native Purification Protocol. Lysate (2 ml, 2 CV) from a 100 ml *E. coli* culture was loaded onto a 1 ml IMAC column. The column was washed with 6 CV of wash buffer 1, followed by 6 CV of wash buffer 2, and purified protein was eluted with 5 CV of elution buffer (all at 2 ml/min). The purified product was >95% pure by densitometric scanning and Quantity One Software analysis. Lane 1, Precision Plus Protein Unstained Standards; lane 2, soluble lysate; lane 3, flowthrough; lane 4, wash 1; lane 5, wash 2; lane 6, purified product.
* Nif-3 construct was kindly provided by R. Stevens, Scripps Institute.

Section 7
Scaling Up

EconoFit Columns are available in 1 and 5 ml formats. Profinity IMAC Resin is also available in various amounts, from 25 ml bottles to larger bulk quantities, for scaling up methods developed using the columns. For quick scale-up, two or three columns of the same type can be connected in series, so take care to maintain an overall system pressure ≤72 psi.

In addition, Bio-Rad carries an extensive line of empty chromatography columns from laboratory to process scale. Ask your local Bio-Rad representative or go to **bio-rad.com/ResinsandColumns** for more information.

Section 8
Regenerating, Cleaning, Sanitizing, and Storing Columns

Protein cross-contamination, frit clogging, and increased backpressure can result from running a column beyond the recommended number of uses. After repeated use, a column may run slower or produce high backpressure, a result that should be expected due to the nature of the sample mixture. The following cleaning and regeneration procedures may be used. However, we recommend that you dispose of a column after several uses. To avoid cross-contamination, we recommend that each column be designated for a single protein. To maintain good flow properties, we recommend that the columns be cleaned between uses.

Run the cleaning protocol at 2 ml/min for 1 ml columns. For 5 ml columns, run the cleaning protocol at 5 ml/min.

High Salt/Acid Cleaning

1. Rinse the column with 2 CV of water at 2 ml/min.
2. Wash the column with 5 CV of 500 mM NaCl, 50 mM Tris, pH 8.0 at 2 ml/min.
3. Wash the column with 5 CV of 500 mM NaCl, 100 mM NaOAc, pH 4.5 at 2 ml/min.
4. Rinse the column with 2 CV of water at 2 ml/min.
5. Store the column in 20% ethanol at 4°C.

Chaotropic Cleaning

1. Rinse the column with 2 CV of water at 2 ml/min.
2. Wash the column with 5 CV of 6 M guanidine HCl at 2 ml/min.
3. Rinse the column with 2 CV of water at 2 ml/min.
4. Store the column in 20% ethanol at 4°C.

Other Cleaning Guidelines

In situations where it is desirable to run different proteins over the same column, completely sanitize, strip, and recharge the column between sample runs. Take care when handling and disposing of metal-containing solutions.

1. Clean the column with 10 CV of 1.0 M NaOH.
2. Rinse the column with 10 CV of water.
3. Strip metal ions with 5 CV of 0.1 M EDTA.
4. Rinse the column with 10 CV of water.
5. Recharge the column with 5 CV of 0.1 M nickel sulfate, pH 4.5.
6. Rinse the column with 10 CV of water.
7. Store the column in 20% ethanol.

Section 9
Troubleshooting Guide

Possible Causes	Possible Solutions
Column Clogging or Slow Flow Rate	
Particulates in sample	Filter all samples and buffers through 0.2 µm filter prior to application
Sample too viscous	Add nuclease to lysate to degrade DNA. Centrifuge and filter lysate again
No Target Protein in Eluate	
Low level of target	Check expression level of protein in starting SDS-PAGE material
Target protein not binding	▪ Check levels of target protein in lysate, flowthrough, wash fractions, and eluted fractions ▪ Check for presence of histidine tag with antihistidine antibody
Target Protein in Flowthrough	
Histidine tag not accessible	▪ Purify protein under denaturing conditions to expose histidine tag ▪ Reclone histidine tag onto opposite terminus (N– or C– terminus)
Proteolysis and removal	Include protease inhibitors in histidine-tagged lysis buffer or purify in the cold
Precipitation during Purification	
Binding capacity of column exceeded	Load less sample
Protein aggregating	▪ Include low amount of detergent (0.1% Triton X-100, Tween 20) ▪ Include glycerol up to 10%
Protein too concentrated in step elution	Elute with imidazole gradient
Eluted Protein Is Impure	
Contaminants coeluting	▪ Elute with imidazole gradient (10–500 mM) rather than step elution ▪ Increase imidazole in the wash to increase wash stringency, but keep below 40 mM
Target Protein Is Degraded	
Proteolysis of target	▪ Add protease inhibitors to protein lysate ▪ Purify at 4°C or under denaturing conditions

Section 10
Ordering Information

Catalog #	Description
EconoFit Profinity IMAC Columns	
12009298	**EconoFit Profinity IMAC Columns**, 5 x 1 ml columns
12009299	**EconoFit Profinity IMAC Column**, 1 x 5 ml column
12009300	**EconoFit Profinity IMAC Columns**, 5 x 5 ml columns
Profinity IMAC, Ni-charged, Resins	
1560131	**Profinity IMAC Resin**, Ni-charged, 10 ml bottle
1560133	**Profinity IMAC Resin**, Ni-charged, 25 ml bottle
1560135	**Profinity IMAC Resin**, Ni-charged, 100 ml bottle
156-0137	**Profinity IMAC Resin**, Ni-charged, 500 ml bottle
Profinity IMAC, uncharged, Resins	
1560121	**Profinity IMAC Resin**, uncharged, 10 ml bottle
1560123	**Profinity IMAC Resin**, uncharged, 50 ml bottle
156-0127	**Profinity IMAC Resin**, uncharged, 1 L bottle

Section 11
Bibliography

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