### **Gene Information**

| Gene Name            | odz, odd Oz/ten-m homolog 1(Drosophila)   |
|----------------------|---|
| Gene Symbol          | ODZ1  |
| Organism             | Human   |
| Gene Summary         | The protein encoded by this gene belongs to the tenascin family and teneurin subfamily. It is expressed in the neurons and may function as a cellular signal transducer. Several alternatively spliced transcript variants encoding different isoforms have been found for this gene. |
| Gene Aliases         | ODZ3, TEN-M1, TNM, TNM1   |
| RefSeq Accession No. | NC_000023.10, NT_011786.16, NG_013249.1   |
| UniGene ID           | Hs.727607   |
| Ensembl Gene ID      | ENSG00000009694   |
| Entrez Gene ID       | 10178   |

## **Assay Information**

| Unique Assay ID               | qHsaCID0015682  |
|-------------------------------|---|
| Assay Type                    | SYBR® Green   |
| Detected Coding Transcript(s) | ENST00000371130, ENST00000422452  |
| Amplicon Context Sequence     | TGATAGATAGAGTGATTCAGACACAGGGTCCATAGCCAGATAGTATTTGTGAGCA<br>GGACTTGTGCTATGTCTGGTATCTCTGTTTCTTAATTCCAAAATACTAACGGAGTT<br>TCCCGAGGGAAATATTCTCCTTACAAAATTGAAGTCGCCAACATACACACTGCC |
| Amplicon Length (bp)          | 114   |
| Chromosome Location           | X:123615751-123630945   |
| Assay Design                  | Intron-spanning   |
| Purification                  | Desalted  |

### Validation Results

| Efficiency (%)    | 98     |
|-------------------|--------|
| R <sup>2</sup>    | 0.9992 |
| cDNA Cq           | 23.46  |
| cDNA Tm (Celsius) | 80.5   |
| gDNA Cq           | 44.28  |



| Specificity (%)  | 100 |
|------------------|-----|
| opecinicity (70) | 100 |

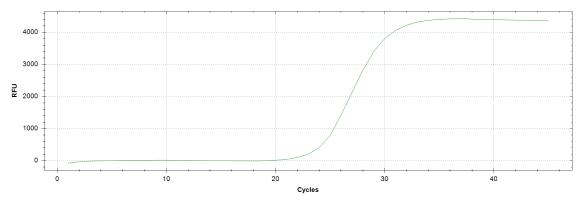
Information to assist with data interpretation is provided at the end of this report.



### ODZ1, Human

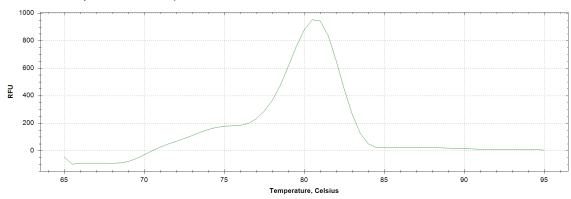
### **Amplification Plot**

Amplification of cDNA generated from 25 ng of universal reference RNA



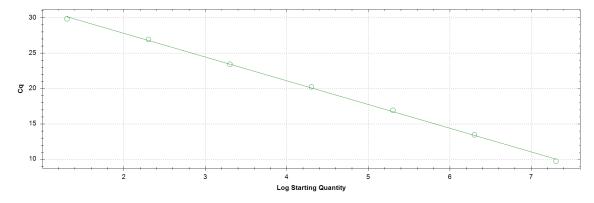
#### **Melt Peak**

Melt curve analysis of above amplification



#### **Standard Curve**

Standard curve generated using 20 million copies of template diluted 10-fold to 20 copies





## Products used to generate validation data

| Real-Time PCR Instrument      | CFX384 Real-Time PCR Detection System            |
|-------------------------------|--|
| Reverse Transcription Reagent | iScript™ Advanced cDNA Synthesis Kit for RT-qPCR |
| Real-Time PCR Supermix        | SsoAdvanced™ SYBR® Green Supermix                |
| Experimental Sample           | qPCR Human Reference Total RNA                   |

### Data Interpretation

| Unique Assay ID               | This is a unique identifier that can be used to identify the assay in the literature and online.  |
|-------------------------------|---|
| Detected Coding Transcript(s) | This is a list of the Ensembl transcript ID(s) that this assay will detect. Details for each transcript can be found on the Ensembl website at www.ensembl.org.   |
| Amplicon Context Sequence     | This is the amplicon sequence with additional base pairs added to the beginning and/or end of the sequence. This is in accordance with the minimum information for the publication of real-time quantitative PCR experiments (MIQE) guidelines. For details, please refer to the following publication, "Primer Sequence Disclosure: A Clarification of the MIQE Guidelines" (Bustin et al 2011). |
| Chromosome Location           | This is the chromosomal location of the amplicon context sequence within the genome.  |
| Assay Design                  | Exonic: Primers sit within the same exon in the mRNA transcript and can potentially co-amplify genomic DNA. If performing gene expression analysis, it is suggested that the samples be treated with a DNase to eliminate potential unwanted signal from contaminating genomic DNA.   |
|                               | Exon-exon junction: One primer sits on an exon-exon junction in mRNA. When performing gene expression analysis, this design approach will prevent unwanted signal from contaminating genomic DNA.   |
|                               | Intron-spanning: Primers sit within different exons while spanning a large intron in the mRNA (intron is greater than 750bp). When performing gene expression analysis, this design approach should limit potential unwanted signal from contaminating genomic DNA.   |
|                               | Small intron-spanning: Primers sit within different exons with a short intron in between (intron is smaller than 750bp). Small introns may not prevent unwanted signal from contaminating genomic DNA.  |
| Efficiency                    | Assay efficiency was determined using a seven-point standard curve from 20 copies to 20 million copies. While an efficiency of 100% represents a perfect doubling of template at every cycle and is ideal, typical ranges of good assay efficiency are between 90-110%. For difficult targets, assay efficiency outside of this range are accepted and reported accordingly.                      |
| R <sup>2</sup>                | The R <sup>2</sup> represents the linearity of the standard curve and how well the standard curve data points fit the linear regression line. Acceptable values are >0.98.  |



| cDNA Cq     | Cq value obtained from 25ng of cDNA transcribed from universal RNA when performing wet-lab validation of the assay.   |
|-------------|---|
|             | Note: Not all genes will be expressed at a detectable level in the universal RNA sample.  |
| cDNA Tm     | Melting temperature of the amplicon when running a melt curve analysis.   |
| gDNA Cq     | Cq value obtained when running the assay with 2.5ng of genomic DNA. This is more than a moderate level of genomic DNA contamination. Intron-spanning and exon-exon junction assay designs can minimize or eliminate genomic DNA detection.  Note: Genomic DNA contamination is often present at variable levels. If concerned |
|             | about genomic DNA contamination, the genomic DNA contamination control assay is recommended to run with your sample to determine if genomic DNA levels are sufficient to negatively impact results.   |
| Specificity | This value is the percent of specific amplicon reads as measured by next generation sequencing (NGS). While 100% specificity is desirable, small decreases in specificity (<1%) can be due to NGS read errors. More significant reductions are likely due to co-amplification of homologous regions.                          |
|             | Note: Since gene expression can be cell type and condition specific, the exact level and impact of co-amplification in a given sample is impossible to predict. If co-amplification is detected, it should be taken into consideration and reported when analyzing gene expression results.                                   |

